Core J Procedure Protocol 10% NCLPPS, Compactin, Kdo2 Lipid A Treatment

Setup

- 1. See Figure 1 for an overview of the treatment time course.
- 2. See Table 1 for an overview of the four treatment groups.
- 3. Maintain sterile technique throughout the treatment procedure until harvesting.
- 4. Plate 2×10^6 cells per 60 mm plate in 5 ml of 10% NCLPPS medium (PS0000002100) as recommended in the LIPID MAPS Thawing and Passage Procedure (PP0000000101). Make triplicate plates for each condition at each time point. Assign a barcode to each plate and enter into LIMS.
- 5. Incubate 24 hours at 37°C.

Reagent Preparation

- 1. Mevalonate 20 mM prepare from 0.2 M mevalonate (PS0000002200)
- 2. Compactin $10 \text{ mM} \text{thaw from } -80^{\circ}\text{C}$
- 3. Kdo2 Lipid A working solution prepare from Kdo2 Lipid A stock solution (PS0000001401)
- 4. Spray the reagent tubes with 70% ethanol and let air dry before using.

Treatment

- 1. Remove 12 dishes from the incubator and label 24A through 24L.
- 2. Before treating the cells, remove 0.5 mL of medium from each plate for the TNFα assay, place in labeled Eppendorf tubes, and place at 4°C until further processing.
- 3. Treat each dish with the appropriate reagents:

Group 1: -C-K	Dishes A, B, C	11.25 μL mevalonate
		22.5 µL PBS
		4.5 µL PBS
Group 2: +C-K	Dishes D, E, F	11.25 μL mevalonate
		22.5 μL compactin
		4.5 PBS
Group 3: -C+K	Dishes G, H, I	11.25 μL mevalonate
		22.5 µL PBS
		4.5 μL Kdo2 Lipid A
Group 4: +C+K	Dishes J, K, L	11.25 μL mevalonate
		22.5 μL compactin
		4.5 μL Kdo2 Lipid A

- 4. Note the time and return the dishes to the incubator.
- 5. Repeat this treatment procedure for every time point, working backwards from 24 hours to 12, 8, 4, 2, 1, and 0.5.
- 6. For 0 hours, do not treat with anything, not even mevalonate. Label three dishes A, B, and C, take the medium for $TNF\alpha$, and proceed with harvesting.

Harvest

- 1. At the appropriate time, remove the dishes from the incubator and place on ice.
- 2. Before harvesting the cells, remove 0.5 mL of medium from each plate for the TNFα assay, place in labeled Eppendorf tubes, and place at 4°C until processing.
- 3. Transfer remaining medium for Core G to a 15 mL conical polypropylene tube on ice.
- 4. Gently wash each plate twice with 3 ml of cold PBS.
- 5. Add 2 mL of PBS/1mM EDTA to each dish and scrape the cells with a scraper.
- 6. Transfer the cells to a 15 mL conical polypropylene tube and pipette 20x with a p1000 to suspend the cells.
- 7. Remove 400 μ L of the cell suspension to an eppendorf on ice for DNA assay.
- 8. Proceed with sterol extraction using the remaining 1.6 mL of cells in PBS.

Medium Preparation for Core G

- 1. Add 100 μ L of Core G Eicosanoid Internal Standard to each medium sample, vortex.
- 2. Remove stray cells by centrifuging the samples at 2400 rpm (1160 rcf) for 5 min at 4°C. (eppendorf 5810 R with swinging bucket rotor)
- 3. Decant medium into a 5 mL polypropylene snap-cap tube. Store at -20°C.
- 4. Send frozen samples on dry ice to Core G. Include a sample of 4 mL medium spiked with 100 μL Eicosanoid Internal Standard, as well as all remaining internal standard mix.

Medium Processing for TNFa Assay

- Centrifuge the TNFα aliquots at top speed in coldroom microfuge for 2 min, collect an aliquot of supernatant (~0.3-0.4 ml) and place in a new labeled Eppendorf tube. Assign a barcode, enter into LIMS and freeze the aliquots at -80°C.
- 2. The TNFα aliquots from the Kdo₂-lipid A treated cells must be diluted in medium at least 1:80 before assaying. ElisaTech will dilute the samples, if requested. Do not dilute aliquots from cells that were not treated with Kdo₂-lipid A.
- Send frozen labeled aliquots to ElisaTech, 12635 E. Montview Blvd., Suite 215, Aurora, CO 80010. For assaying in house, use the Quantikine mouse TNFα/TNFSF1A EIA kit (R&D Systems, Cat. #MTA00).

Cell Processing for DNA Assay

- 1. Add 20 μ L of 50% etOH in H₂O to each sample for DNA analysis. Store at -20°C.
- Assign a barcode and enter into LIMS. Follow the DNA assay protocol in Molecular Probe's manual with two exceptions: 1) Use 5 ul of standards instead of 10 ul; and 2) Substitute component B with Molecular Probes' Lysis Buffer (cat C-7027) at 1x concentration.

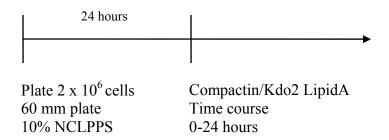


Table 1

	Group 1: -C-K	Group 2: +C-K	Group 3: -C+K	Group 4: +C+K
Mevalonate 50 µM	+	+	+	+
Compactin 50 µM	-	+	-	+
Kdo2 LipidA 100 ng/mL	-	-	+	+

Author: Bonne Thompson Date: 3/13/06